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GRANT NUMBER DAMD17-94-J-4386

TITLE: Detection and Characterization of Autoantigens in
Breast Cancer

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REPORT DATE: August 1997

TYPE OF REPORT: Annual

PREPARED FOR: Commander
U.S. Army Medical Research and Materiel Command
Fort Detrick, Frederick, Maryland 21702-5012

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DTIC QUALITY INSPECTED 3

19980114 102

REPORT DOCUMENTATION PAGE			Form Approved OMB No. 0704-0188 RACEVSKIS, Janis	
Public reporting burden for this collection of information is estimated to average 1 hour per response, including the time for reviewing instructions, searching existing data sources, gathering and maintaining the data needed, and completing and reviewing the collection of information. Send comments regarding this burden estimate or any other aspect of this collection of information, including suggestions for reducing this burden, to Washington Headquarters Services, Directorate for Information Operations and Reports, 1215 Jefferson Davis Highway, Suite 1204, Arlington, VA 22202-4302, and to the Office of Management and Budget, Paperwork Reduction Project (0704-0188), Washington, DC 20503.				
1. AGENCY USE ONLY (Leave blank)	2. REPORT DATE August 1997	3. REPORT TYPE AND DATES COVERED Annual (15 Jul 96 - 14 Jul 97)		
4. TITLE AND SUBTITLE Detection and Characterization of Autoantigens in Breast Cancer		5. FUNDING NUMBERS DAMD17-94-J-4386		
6. AUTHOR(S) Janis Racevskis, Ph.D.				
7. PERFORMING ORGANIZATION NAME(S) AND ADDRESS(ES) Montefiore Medical Center Bronx, New York 10467		8. PERFORMING ORGANIZATION REPORT NUMBER		
9. SPONSORING/MONITORING AGENCY NAME(S) AND ADDRESS(ES) Commander U.S. Army Medical Research and Materiel Command Fort Detrick, Frederick, MD 21702-5012		10. SPONSORING/MONITORING AGENCY REPORT NUMBER		
11. SUPPLEMENTARY NOTES				
12a. DISTRIBUTION / AVAILABILITY STATEMENT Approved for public release; distribution unlimited			12b. DISTRIBUTION CODE	
13. ABSTRACT (Maximum 200) The entire sequence of our second breast tumor associated autoantigen (Auag2), which is a newly discovered gene product, has been determined. The predicted amino acid sequence of Auag2 contains two LIM domain motifs, which are conserved cysteine rich zinc-binding motifs of about 60 amino acid residues that mediate protein-protein interactions, and are characteristic of a group of critical transcriptional regulators of embryonic development. The 350 bp region coding for the two tandem LIM domains was found to be 60% homologous to the analogous region of rhombotin 1, a proto-oncogene of 160 amino acids whose gene is disrupted by chromosomal translocation in T-cell leukemia. A third autoantigen clone has been identified and is being characterized. Various features of the Auag2 cDNA sequence (a long GC-rich structured 5' end, the presence of mRNA destabilizing motifs in the 3' end and a homology to a known oncogene) all predict that this gene plays a vital role in the life of the organism, and may play a role in malignancy.				
14. SUBJECT TERMS Breast Cancer			15. NUMBER OF PAGES 14	
			16. PRICE CODE	
17. SECURITY CLASSIFICATION OF REPORT Unclassified	18. SECURITY CLASSIFICATION OF THIS PAGE Unclassified	19. SECURITY CLASSIFICATION OF ABSTRACT Unclassified	20. LIMITATION OF ABSTRACT Unlimited	

FOREWORD

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Jarin Racunski 9/9/97
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INTRODUCTION

Tumor growth is associated with the expression of mutated gene products, inappropriate gene expression, and the breakdown of tissue architecture, leading to the exposure and release into the peripheral circulation of sequestered antigens (1,2). Whether these circulating, mutated or newly displayed tumor-associated antigens elicit an autologous humoral immune response in the breast tumor patient is of vital interest. Isolation, identification and characterization of novel breast tumor associated autoantigens might yield new insights into the disease process, and moreover, may be developed into diagnostic screening tests and potential targets for immunotherapy.

The screening of cDNA expression libraries with autologous patient serum is a powerful technique, which has been used successfully for the identification of autoimmune disease antigens (3), and which we have adapted for the identification of autoantigens in cDNA libraries made from breast tumor mRNA. After screening cDNA libraries, derived from primary ductal breast carcinomas with autologous patient serum, we have detected and isolated three immunoreactive cDNA clones, all three of which are newly discovered gene products. The first autoantigen isolate *Ngp 1* has been characterized and is a nucleolar GTP-binding protein. The predicted amino acid sequence of the second clone *Auag2* contains two LIM domain motifs and bears a 60% homology in this region to a known oncogene, Rhombotin 1. Sequencing of the third isolate *Auag3* is underway. Our studies have identified novel proteins that might be involved in malignancy, and may help understand the biology of breast cancer.

BODY

Most of our effort for the past year has focused on characterizing our second breast tumor autoantigen isolate (working name *Auag2*), which is also a newly discovered gene. We chose to focus on *Auag2* because we found it to be related to a group of known oncogenes, and it may represent an important new discovery. Work is continuing on our first autoantigen isolate *Ngp-1*, the nucleolar GTP-binding protein (4), to identify other proteins that interact with it using the yeast two-hybrid vector system (5). We have encountered difficulties using the original phagemid vector *pBD-GAL4* as the bait plasmid expressing the open reading frame of *Ngp-1*, and will continue this work using recently available more tightly controlled inducible vectors (*pGILDA*). The inducible vectors allow performing two-hybrid library screening with bait proteins that interact with yeast host cell proteins, or are toxic to yeast cells. The highly conserved nature of *Ngp-1* which we even found to have significant homology to a gene from such a distant organism as rice (4), probably accounts for the difficulties we encountered. We have isolated a third autoantigenic clone fragment (*Auag3*) which, with the exception of expressed sequence tags in the databases, shows no homology to any known gene (Figure 5, page 14). A 3kb clone containing *Auag3* sequences has been isolated and is being characterized.

Auag2 cDNA contains a long region of extremely high GC content in the 5' third of the molecule (Figure 1, page 11) which interferes with reverse transcription, hence full length clones of *Auag2* are under-represented in cDNA libraries. This was illustrated by our cDNA homology searches of the databases, which identified a number of identical human expressed sequence tags, however, none of them representing the first 900 bases of the *Auag2* cDNA. Furthermore, we found that standard PCR reactions could not amplify the GC rich region, and special formulations which melt high GC DNA had to be employed. A full length clone containing 2.1 kb of *Auag2* sequence was isolated by using the Gene Trapper technology, where a cDNA library in a plasmid vector is converted to single strand form, then hybridized with gene specific biotinylated oligonucleotide probes which are captured by avidin coated magnetic beads, thus highly enriching for the desired gene product. The high GC content also made accurate sequencing a difficult task and required the use of dITP in the sequencing reactions and formamide sequencing gels. Clones of *Auag2* have been isolated from two different cDNA libraries (brain and testis), and the entire sequence determined accurately (*Genbank accession # U24576*). The first ATG codon in the preferred configuration with an A in position -3, and a G in position +4 (6), is located at position 781 at the start of an open reading frame which could encode a protein of 165 amino acids (Figure 1, page 11). The predicted amino acid sequence of *Auag2* contains two LIM domain motifs (Figure 2, page 12), which are conserved cysteine rich zinc-binding motifs of about 60 amino acid residues that mediate protein-protein interactions, and are

characteristic of a group of critical transcriptional regulators of embryonic development (7). A search of the databases revealed that the 350 bp region coding for the two tandem LIM domains was 60% homologous to the analogous region of rhombotin 1 (Figure 3, page 13), a proto-oncogene of 160 amino acids whose gene is disrupted by chromosomal translocation in T-cell leukemia (8,9). At the amino acid level, the homology, and spacing of the amino acids making up the LIM domains is even more apparent (Figure 4, page 13). These sequences are so highly conserved that the homology to the rhombotin 1 homologue of *Drosophila* is just as extensive (Figure 4, page 13). A number of the expressed sequence tags found in homology searches were from mouse cDNA libraries and showed a greater than 97% homology, also indicating that *Auag2* is highly conserved in evolution. Although the long untranslated 5' end of *Auag2* shows a slight homology to some other genes with GC rich regions such as vascular endothelial growth factor, the sequence is unique, as is the long 3' end. *Auag2* does not seem to have any other closely related genes, since after extensive screening of different cDNA libraries with cDNA probes, in our efforts to obtain full length clones, we have not isolated any related cDNAs. We did however isolate one 5' end splice variant numerous times. The most extensively studied of the rhombotins (three known members) is rhombotin 2, which has also been shown to be a proto-oncogene in T-cells, and act as a transcriptional regulator of erythroid development (9,10). Rhombotin 2 is widely distributed in many tissues with the exception of T-lymphocytes, and is believed to have multiple, as yet unknown regulatory functions. As judged by northern blot analysis, the distribution of *Auag2* expression is unlike that of RBTN1 or RBTN2. In addition, the long, unique, 5' and 3' regions of *Auag2* suggest that *Auag2* has different control elements specifying other tissue distribution and regulatory functions.

The extremely long 5' end of *Auag2* (780 bp) is the GC rich region (Figure 1, page 11). A long GC rich, structured 5'-leader sequence is characteristic of transcripts encoding oncoproteins, growth factors, transcription factors, and other regulatory proteins - that seem to be designed to be translated poorly (11). Inhibition at the translational level seems to be a component of gene regulation for genes which need to be tightly regulated. Another feature of the sequence of *Auag2* cDNA is the presence of multiple ATTT motifs in the 3' end (Figure 1), which have also been observed in the 3' untranslated region of numerous lymphokine, cytokine, and proto-oncogene mRNAs. It has been proposed that such ATTT motifs are involved in the selective degradation of transiently expressed messengers (12). In northern blots of mRNAs isolated from various human tissues, *Auag2* appears to be most highly expressed in testes and brain, and is not detectable in liver and kidney. *Auag2* transcripts are also present in mRNA extracted from breast tumors, however, since breast tumors are a complex mixture of different cell types (stromal fibroblasts, infiltrating lymphocytes and transformed breast epithelial cells), we will have to do immunohistochemistry and in-situ hybridization to determine the

exact source of these transcripts. In certain tissues there appears to be an extra band; probably representing different splicing products. A splice variant involving the 5' end of *Auag2* cDNA has been isolated from different cDNA libraries; its role in the expression of *Auag2* protein is yet to be determined. Because of the highly conserved nature of the *Auag2* protein, it may prove to be a weak immunogen in rabbits. An alternative is to use the anti-peptide antibody approach which enables the animal host to bypass the immune tolerance frequently encountered when a highly conserved or self protein antigen is used as the immunogen. We therefore have obtained antiserum against a synthetic peptide from the open reading frame amino terminus region, outside the LIM domain, which has no homology to the rhombotins, for immunohistochemical localization of *Auag2* protein within different tissues and cell types.

Since LIM domain containing proteins interact with other proteins to form specific transcription regulators, we will try to identify those that interact with *Auag2* by applying the yeast two hybrid system. Rhombotin 2, for example, has been shown to interact with retinoblastoma-binding protein 2 (10).

CONCLUSIONS

Clones of *Auag2* have been isolated from two different cDNA libraries and the entire sequence determined accurately. The predicted amino acid sequence of *Auag2* contains two LIM domain motifs, which are conserved cysteine rich zinc-binding motifs of about 60 amino acid residues that mediate protein-protein interactions, and are characteristic of a group of critical transcriptional regulators of embryonic development. A search of the databases revealed that the 350 bp region coding for the two tandem LIM domains was 60% homologous to the analogous region of rhombotin 1, a proto-oncogene of 160 amino acids whose gene is disrupted by chromosomal translocation in T-cell leukemia. As judged by northern blot analysis, the distribution of *Auag2* expression is unlike that of *RBTN1* or *RBTN2*. In addition, the long, unique, 5' and 3' regions of *Auag2* suggest that *Auag2* has different control elements specifying other tissue distribution and regulatory functions.

Various features of the *Auag2* cDNA sequence (a long GC-rich structured 5' end, the presence of mRNA destabilizing motifs in the 3' end and a predicted amino acid sequence which contains two LIM domain motifs with a partial homology to a known oncogene) all predict that this gene plays a vital role in the life of the organism, and merits further investigation and characterization.

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1   GCTCTGTCAGTAACACATGTGTAAGAGCCGCGGAGGGAGCGAGCGAGCCGGCTAGAGGCC
61  AGCGCGCGCGCGCGCGCGCGCTCCGAGCCGGGCAGCAACAGCCCCGGCAGCGGCGCAGGC
121 TCCAGCGCGCGCGCGCGCGCGCGCAGCCCCGACGCGCTGGGTGCGCCTGCCCTGCCGG
181 CGTCCGCACCGTCCGCGCGCGCTCCCGGGGTGTTGTGTCTGGAGTGGTCCCCGGCCGGA
241 GGTGCAGGGAGCTCAGCCGAGCCGCGCTGCCATCCCGGAGCGAGCAAGCGAGCGAGCGC
301 GCGGGAGGGAGGAAGGCGGCGGCGGAGGAGGAGGAGCGGGAGGAGCGCGGGCGGGGG
361 CGGGGGCGCGCGCGGGGGAATATACAAAGTGAAGCCACATTGCCAAACTTGACAGCAGC
421 GATTGCAGCAGTTGCTGCGCTGCGCGCGCGCTGAAGCCGCGCGCGCGGGCGGAGGGCT
481 CCTGCAGCTGCTCGCGCGCAGTCCGAGGCGGAGAAGGACGAAGACTGAGACTGACACTTC
541 TGCTCCCGCGCGCGCGCGCACTTACGCGGGGGCCCCCAACCGCCCCAGAGCAACCGGAT
601 TTAACAAAAAAGCCGCGCTTAGCCCCCTCCCTCCCTTTCTGCTTCTGCGAGAA
661 CTCCCTCCCTCCCTCCAGCTCCGCGAGCGCAGCGCGCGCTTCCCTGGAAGCGGAGCGGCT
721 TCGCTCGCATTTCACCGCGCGCGCTCTCGCAATATTGCAATATAGGGGAAAAGCAGACC
781 ATGGTGAATCCGGGCAGCAGCTCGCAGCCGCCCCCGGTGACGGCCGGCTCCCTCTCCTGG
841 AAGCGGTGCGCAGGCTGCGGGGGCAAGATTGCGGACCGCTTTCTGCTCTATGCCATGGAC
901 AGCTATTGGCACAGCCGGTGCCCTCAAGTGCTCCTGCTGCCAGGCGCAGCTGGGCGACATC
961 GGCACGTCTGTACACCAAAAGTGGCATGATCCTTTGCAGAAATGACTACATTAGGTTA
1021 TTTGGAAATAGCGGTGCTTGCAGCGCTTGCAGCAGTTCCTGCGAGTGAACCTGTC
1081 ATGAGGGCGCAAGGCAATGTGTATCATCTTAAGTGTTTTACATGCTCTACCTGCCGAAT
1141 CGCCTGGTCCCAGGAGATCGGTTTCACTACATCAATGGCAGTTTATTTTGTGAACATGAT
1201 AGACCTACAGCTCTCATCAATGGCCATTTGAATTCACCTCAGAGCAATCCACTACTGCCA
1261 GACCAGAAGGTCTGCTTAAAAGGTGAGTAATGCAGAATGCGTGCCCTTCATCTCAGATTT
1321 GTTCATCACAGGTGGATCCCATGTGTCTTCAGTAGACAAGTCACCTTTGTAGCTAGCACC
1381 AGTGCCAGCTCCATGCCATTGCACCTTCTTTAGTCTTGATTGCCCTTCCCGCATTATTTG
1441 GTGTATTAAATGACTGAATATGAACATTAAGGACTCCATGAACCTGGGCTAATGGGAGA
1501 CTGTAGAGAAAATGAAAAAGATCCACCAGAGGACATCTTGGGGAGGGGGAGGGAGCTGG
1561 GGGGGAGGGAAATGACTAATGAAGCTAATTAAGAAGCATTCAAATCTGCTTTCTACCC
1621 TCATTAACAATTAGCAGGGCACTGGCCAGAGTTTGTACCCTGTGTTTTACCTTAACAACA
1681 TTCTATTTGCTCTTTGTATATTTAAGTGTGTGAAGGAAACGTGTTTCAATCAAACTGAC
1741 CATGAGATAAAGGAAAGAGATGTGGCTTTTGTGATATTCTATCACAAACACTTATTGTAT
1801 CTCTGTAAAATACAATGTATGTATGCATGTAAGTGTTTTGTCCATATGTTGCTACTCCC
1861 ATGGCAAAGAAAAAAGAAATGAAAAAAGAAAAAAATTTGGAAAAAATCAGGC
1921 TCATAGCAGCTACTGTGTAGAAAATTCCCCCTACTTCTAATTTGCTGAATGAAGAAAAA
1981 AAAAATCTTTTATTTGTGATATTTTCAGAGACATTTGCTCTAGTATGGTGTATTTAAATA
2041 ATAAAACTTAAAAGAAAAATAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAA
2101 AAAAAAAAAAAAAAAAAAAAAAA 3'

```

Figure 1. Complete nucleotide sequence of *Auag2* cDNA (Genbank accession # U24576). Initiation (nucleotide 781) and termination codons (nucleotide 1278) of the putative open reading frame are shown in bold double underline. G and C residues in the 5' untranslated region (nucleotides 1-780) are shaded, and AATT motifs in the 3' region are underlined. Potential polyadenylation signal **AATAAA** is in bold at position 2040.

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1   M V N P G S S S Q P P P V T A G S L S W K
781 ATGGTGAATCCGGGCAGCAGCTCGCAGCCGCCCCGGTGACGGCCGGCTCCCTCTCCTGGAAG

22  R C A G C G G K I A D R F L L Y A M D S Y
844 CGGTGCGCAGGCTGCGGGGGCAAGATTGCGGACCGCTTTCTGCTCTATGCCATGGACAGCTAT

43  W H S R C L K C S C C Q A Q L G D I G T S
907 TGGCACAGCCGGTGCTCAAGTGCTCCTGCTGCCAGGCGCAGCTGGGCGACATCGGCACGTCC

64  C Y T K S G M I L C R N D Y I R L F G N S
970 TGTTACACCAAAGTGGCATGATCCTTTGCAGAAATGACTACATTAGGTTATTTGGAAATAGC

85  G A C S A C G Q S I P A S E L V M R A Q G
1033 GGTGCTTGCGAGCGCTTGCGGACAGTCGATTCTGCGAGTGAACTCGTCATGAGGGCGCAAGGC

106 N V Y H L K C F T C S T C R N R L V P G D
1096 AATGTGTATCATCTTAAGTGTTTTACATGCTCTACCTGCCGGAATCGCCTGGTCCCGGGAGAT

127 R F H Y I N G S L F C E H D R P T A L I N
1159 CGGTTTCACTACATCAATGGCAGTTTATTTGTGAACATGATAGACCTACAGCTCTCATCAAT

148 G H L N S L Q S N P L L P D Q K V C *
1222 GGCCATTTGAATTCACTTCAGAGCAATCCACTACTGCCAGACCAGAAGGTCTGCTAA

```

Figure 2. Predicted amino acid sequence of the *Auag2* open reading frame. LIM domain motif amino acids are shaded and conform to the consensus sequence of all known LIM domains (7):

$[C X_2 C \dots X_{16-23} \dots H X_2 C X_2 C X_2 C \dots X_{16-21} \dots C X_{2-3} (C, H, D)]$

	850	860	870	880	890	900
AUAG2.DNA	AAGCGGTGCGCAGGCTGCGGGGCAAGATTGCGGACCGCTTTCTGCTCTATGCCATGGAC					
	::: : :: :: X::::	::::::::::	::::::::::	::::::::::	: :: :::::	
RBTN1.DNA	AAGGGCTGTGCGGGCTGTAACCGCAAGATCAAGGACCGCTATCTGCTGAAGGCATTGGAC					
	570	580	590	600	610	620
	910	920	930	940	950	960
AUAG2.DNA	AGCTATTGGCACAGCCGGTGCCTCAAGTGCTCCTGCTGCCAGGCGCAGCTGGGCGACATC					
	: :: :::::	::::::::::	: :::::	: :::::	: :::::	:
RBTN1.DNA	AAGTACTGGCACGAAGACTGCCTCAAGTGTCGTGCTGTGACTGCCGCCTGGGCGAGGTG					
	630	640	650	660	670	680
	970	980	990	1000	1010	1020
AUAG2.DNA	GGCACGTCTGTACACCAAAGTGGCATGATCCTTTGCAGAAATGACTACATTAGGTTA					
	::: : :: :::::	: :::::	: :::::	: :::::	: :::::	: :::::
RBTN1.DNA	GGCTCCACCCTCTACACCAAGGCCAACCTCATCCTGTGCCGACGCGACTACCTGAGGCTC					
	690	700	710	720	730	740
	1030	1040	1050	1060	1070	1080
AUAG2.DNA	TTTGGAAATAGCGGTGCTTGCGAGCGCTTGCGGACAGTCGATTCTGCGAGTGAACCTCGTC					
	::::: : : ::	: ::	::::::::::	: ::	: ::	: ::
RBTN1.DNA	TTTGGCACCACAGGGAAGTGTGCTGCTTGCGAGCAAGCTGATCCAGCCTTCGAGATGGTG					
	750	760	770	780	790	800
	1090	1100	1110	1120	1130	1140
AUAG2.DNA	ATGAGGGCGCAAGGCAATGTGTATCATCTTAAGTGTTTTACATGCTCTACCTGCCGGAAT					
	::: ::::: : ::::	::::::::::	: :::::	: :::::	: :::::	: ::X
RBTN1.DNA	ATGCGGGCCCGGACAACGTGTATCACCTCGACTGCTTCGCCTGCCAGCTCTGCAACCAG					
	810	820	830	840	850	860

Figure 3. Alignment of the region encoding the LIM domains of *Auag2* and Rhombotin 1 (RBTN1).

AUAG2	21	KRCAGCGGKIADRFLLYAMDSYWHSRCLKSCCQAQLGDIGTSCYTKSGMILCRNDYIRL	80
		K CAGC KI DR+LL A+D YWH CLKC+CC +LG++G++ YTK+ +ILCR DY+RL	
RBTN1	22	KGCAGCNRKIKDRYLLKALDKYWHEDCLKCACDCRLGEVGSTLYTKANLILCRRDYLR	81
AUAG2	81	FGNSGACSACGQSIPASELVMRAQGNVYHLKCFTCSTCRNRLVPGDRFHYINGSLSFCEHD	140
		FG +G C+AC + IPA E+VMRA+ NVYHL CF C C R GD+F N + C+ D	
RBTN1	82	FGTTGNCAACSKLIPAFEMVMRARNVYHLDCFACQLCNQRFVGDKFFLKNNMILCQMD	141
AUAG2	23	CAGCGGKIADRFLLYAMDSYWHSRCLKSCCQAQLGDIGTSCYTKSGMILCRNDYIRLFG	82
		CAGCG I DR+LL A+D WH CLKC CC +LG++G++ YTK ++LC+ DY+RLFG	
DROS	45	CAGCGKHIQDRYLLRALDMLWHEDCLKCGCCDCRLGEVGSTLYTKGNLMLCKRDYLR	104
AUAG2	83	NSGACSACGQSIPASELVMRAQGNVYHLKCFTCSTCRNRLVPGDRFHYINGSLSFCEHDRP	142
		N+G C+AC + IPA E+VMRA+ NVYHL+CF C C +R GDRF+ + CE+D	
DROS	105	NTGYCAACSKVIPAFEMVMRARTNVYHLECFACQCNHRFCVGDRLFYLCENKILCEYDYE	164

Figure 4. Alignment of the amino acid sequences of the LIM domains of *Auag2* with Rhombotin 1 (RBTN1), and *Drosophila* homologue of RBTN1 (DROS).

```
AUAG3  GGTATCTGGGCAGCCACTGCCAGCATCATCTTTGCTTTCTTGGGATTTGATAACCATTGCT 61
      |||
Z16206 GGTATCTGGGCAGCCACTGCCAGCATCATCTTTGCTTTCTTGGGATTTGATAACCATTGCT 83

AUAG3  GCACCTGCTGCGGAAGTTAAGAATCCACAGAAAACGATGGCACGTGGTATCATCGGGACG 121
      |||
Z16206 GCACATGCTGCGGAAGTTAAGAATCCACAGAAAACGATGGCACGTGGTATCATCGGGACG 143

AUAG3  GTTTTGATTTTCAGCCCTACTCTATGTGTTGTTTCGCCGTTGTTTTGACTGGTATTGTGAAC 181
      |||
Z16206 GTTTTGATTTTCAGCCCTACTCTATGTGTTGTTTCGCCGTTGTTTTGACTGGTATTGTGAAC 203

AUAG3  CTATAAAAAGTTGG 195
      |||
Z16206 TATAAAAAGTTGGG 217
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Figure 5. 5' end sequences of the 3kb isolate of *Auag3* aligned with an expressed sequence tag (Z16206) from a human cDNA library.